

LETTER

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Cerebral spinal fluid biomarker profiles in CNS infection associated with HSV and VZV mimic patterns in Alzheimer's disease

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Alzheimer's disease (AD) is the most common cause of dementia. Although AD was initially considered to be a cell autonomous neurodegenerative disorder, marked neuroinflammation has been observed in brains of AD patients. Genetic and molecular biological findings have suggested the central nervous system (CNS) inflammatory processes to be involved in the etiopathogenesis of AD, in which the activated microglia play a key role. This has also been supported by epidemiological observation that CNS infections are associated with the development of AD, and the relationship between herpes simplex virus (HSV)-1 and AD has been particularly well investigated [1]. For example, the presence of anti-HSV antibody is associated with an elevated risk of AD [2]. Anti-herpetic medication is associated with a reduced risk of dementia in a population-based study [1]. Similar results have also been observed in varicella zoster virus (VZV) infections [3]. In this study, we enrolled 9 patients with HSV infection of the CNS, 8 patients with herpes zoster complicated by CNS involvement, and 18 age-matched control patients presenting with neither CNS infection nor dementia, and measured cerebral spinal fluid (CSF) levels of $A\beta_{1-42}$, $A\beta_{1-40}$, total-tau (t-tau), and phosphorylated tau at threonine 181 (p-tau) as the AD signature; neurofilament light chain (NfL) and phosphorylated neurofilament heavy chain (p-NfH) as indicators of axonal injury; soluble triggering receptor expressed on myeloid cells 2 (sTREM2) as a potential

biomarker for microglial activity; and glial fibrillary acidic protein (GFAP) as a biomarker for astrocytic damage. We also measured serum levels of NfL as a blood-based biomarker for axonal injury. Detailed methods are provided in [Supplementary Methods](#). There was no significant difference in age or sex among the HSV, VZV, and control groups (Table S1). The raw data on biomarkers are presented in Table S2.

We found that the levels of CSF $A\beta_{1-42}$, $A\beta_{1-40}$, and the $A\beta_{1-42}/A\beta_{1-40}$ ratio were significantly lower in the HSV + VZV combined group (HSV/VZV) compared with the control group ($p = 0.01836$, 0.0380 , and 0.0262 , respectively) (Fig. 1a–c). The CSF t-tau, p-tau, sTREM2, and GFAP levels were significantly elevated in the HSV/VZV group compared with the control group ($p = 0.0043$, 0.0007 , 0.0030 , and 0.0139 , respectively) (Fig. 1d, e, i, and j). These results correspond to previous reports showing significantly decreased $A\beta_{1-42}$, increased t-tau, and increased p-tau in CSF of patients with HSV encephalitis [4, 5]. The CSF p-tau/t-tau, CSF NfL, CSF p-NfH and serum NfL levels did not significantly differ between the HSV/VZV and control groups (Fig. 1f, g, h, and k). Comparison among the HSV, VZV and control groups showed that the elevation of CSF p-tau was significant in the VZV group while the level of CSF t-tau was elevated specifically in the HSV group (Supplementary Fig. S1). The other biomarkers showed similar trends to those in comparison between HSV/VZV and the control groups.

Results of uni- and multivariate regression analyses between those biomarker values and clinical severity are summarized in Supplementary Table S3. The negative correlations between Glasgow coma scale and NfL in

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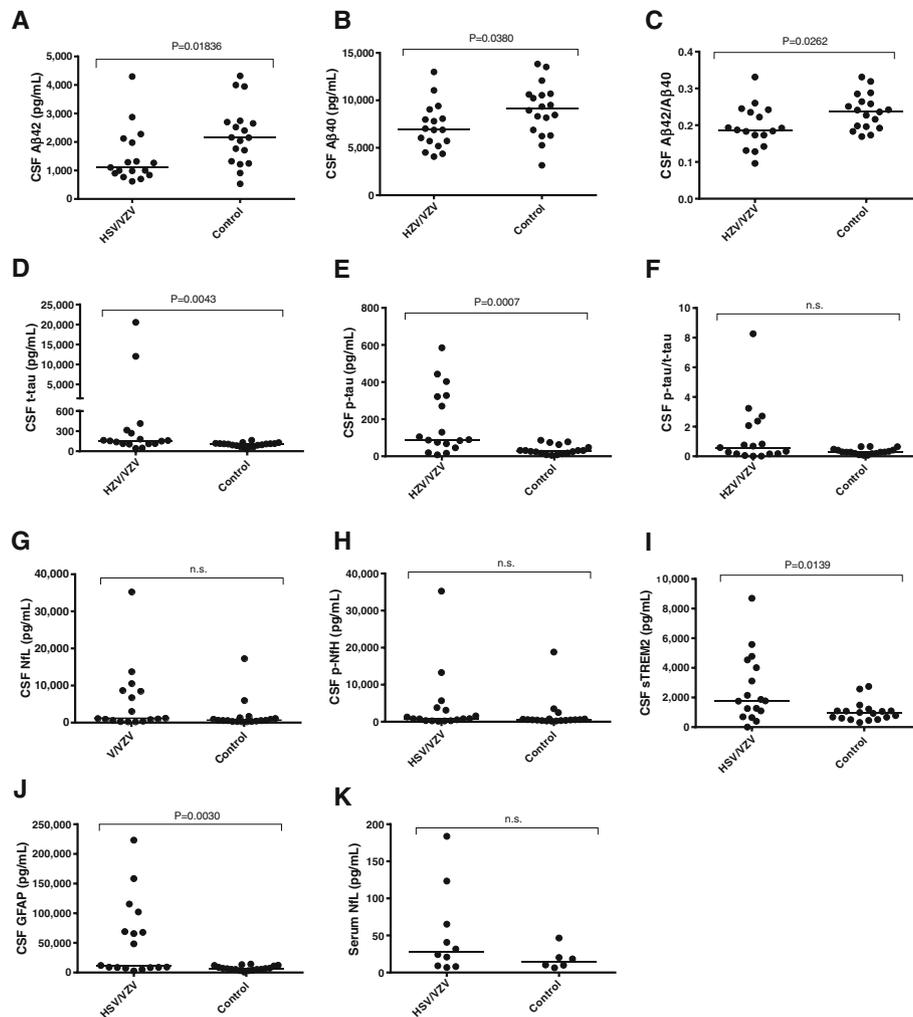


Fig. 1 Comparisons of (a) CSF A β _{1–42}, (b) CSF A β _{1–40}, (c) CSF A β _{1–42}/A β _{1–40}, (d) CSF t-tau, (e) CSF p-tau, (f) CSF p-tau/t-tau ratio, (g) CSF NfL, (h) CSF pNfH, (i) CSF sTREM2, (j) CSF GFAP, and (k) serum NfL between the HSV/VZV and control groups. The CSF (HSV/VZV group: $n = 17$; control: $n = 18$) and serum (HSV/VZV group: $n = 9$; control: $n = 6$) levels of those biomarkers in each individual are shown as black dots. Bars indicate median values. n.s., not significant

CSF and serum were significant after age adjustment ($p = 0.014$ and 0.030 , respectively). Among the biomarkers, only CSF NfL was significantly correlated with the Modified Rankin Scale on discharge after age adjustment ($p = 0.018$). Supplementary Fig. S2 shows scatter plots in cases showing significant correlations on univariate analyses.

The current study has three major implications. First, to our best knowledge, this is the first report of CSF p-tau elevation in patients with CNS VZV infection. This suggests that the biomarker profile of decreased A β _{1–42}, and increased t-tau and p-tau in CSF might be shared not only by CNS involvement of HSV infection but also by CNS VZV infection. This combination of biomarker changes, the so-called “AD

signature”, has been considered to indicate the presence of AD pathology. In particular, high levels of p-tau at threonine181 have been reported to occur solely in AD and not in other neurodegenerative disorders or acute brain damage, such as acute brain infarction [6]. The CSF p-tau elevation in the HSV/VZV group might be attributed to the herpetic infection-induced APP mis-metabolism [7], as similarly seen in the case of AD. In addition, the fact that the biomarker profile in AD patients mimics that in patients with CNS HSV and VZV infections suggests that the latter may be a confounding factor in the CSF biomarker-based diagnosis.

Second, the elevations of CSF sTREM2 and GFAP in the HSV and VZV groups are in line with previous

observations [8]. These trends are also consistent with the reported biomarker changes in patients with AD [9, 10].

Third, only the NfL levels were significantly correlated with the severity and a poor outcome after age adjustment in the CSF biomarkers. This suggests that the NfL concentration in CSF obtained for the diagnostic purpose on admission is the most powerful predictive marker for the severity and prognosis in patients with herpes virus infections among the molecules tested in this study. The CSF and blood NfL levels are reported to be tightly correlated in neurological disorders. Here we found that the serum levels of NfL were strongly associated with their matching CSF levels (Supplementary Fig. S3) and consequently, the ability of serum NfL to evaluate the severity and to predict prognosis may be equivalent to those of CSF NfL.

We acknowledge that the small sample size was a major limitation of this study. Furthermore, the short follow-up period may have weakened the statistical power to detect an association between the prognosis and biomarkers. In the future, it will be necessary to conduct large-scale case-control studies and prospective observations in order to validate the clinical significance of AD-related biomarkers in patients with CNS HSV and VZV infections.

Supplementary Information

The online version contains supplementary material available at <https://doi.org/10.1186/s40035-020-00227-w>.

Additional file 1: Supplementary methods. Table S1: Characteristics of the participants. **Table S2.** The concentrations of biomarkers in participants. **Table S3.** Regression analyses of clinical status and biomarkers. **Figure S1.** Comparison of biomarker concentrations among the HSV, VZV, and control groups (A: CSF A β_{1-42} , B: CSF A β_{1-40} , C: CSF A $\beta_{1-42/1-40}$ ratio, D: CSF t-tau, E: CSF p-tau, F: CSF p-tau/t-tau ratio, G: CSF NfL, H: CSF pNfH, I: CSF sTREM2, J: CSF GFAP, and K: serum NfL). **Figure S2.** Scatter plots of the biomarkers vs the lowest score of GCS during hospitalization (A, B, C, D) as well as the biomarkers vs mRS score at discharge (E, F, G, H, I, J, K) in the HSV/VZV group. **Figure S3.** Correlations between CSF and serum NfL levels. There was a strong positive correlation between CSF and serum NfL levels by Spearman's rank correlation test ($p < 0.0001$).

Abbreviations

CNS: Central nervous system; AD: Alzheimer's disease; HSV: Herpes simplex virus; VZV: Varicella zoster virus; A β : Amyloid β ; t-tau: Total tau; p-tau: Tau phosphorylated at threonine 181; NfL: Neurofilament light chain; p-NfH: Phosphorylated neurofilament heavy chain; GFAP: Glial fibrillary acidic protein; sTREM2: Soluble triggering receptor expressed on myeloid cells 2; CSF: Cerebrospinal fluid

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Authors' contributions

F. K-M., T.O. and Y.F. assisted with patient enrollment, data analysis, and interpretation. H.T., F.K-M., and M.S. performed laboratory work and data analysis. D.A. and T.M. participated in review and revision of the manuscript. M.S., T.K.

and T. T were involved in conceptualization and design of the study, patient enrollment, data collection, interpretation of the data, and review of the manuscript. All authors reviewed the drafts and approved the final version of the manuscript.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article and its supplementary information files.

Ethics approval and consent to participate

Written informed consent from the participants was obtained when possible and, if not, from the nearest relative. The study protocols were approved by the University Ethics Committee (ERB-G-12, Kyoto Prefectural University of Medicine, Kyoto, Japan).

Consent for publication

Not applicable.

Competing interests

The authors had no competing financial interests. Also, no non-financial conflicts of interest existed.

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