

SHORT REPORT

Open Access

In silico analysis of regulatory networks underlines the role of miR-10b-5p and its target *BDNF* in huntington's disease

Sören Müller

Abstract

Non-coding RNAs (ncRNAs) play various roles during central nervous system development. MicroRNAs (miRNAs) are a class of ncRNAs that exert their function together with argonaute proteins by post-transcriptional gene silencing of messenger RNAs (mRNAs). Several studies provide evidence for alterations in miRNA expression in patients with neurodegenerative diseases. Among these is huntington's disease (HD), a dominantly inherited fatal disorder characterized by deregulation of neuronal-specific mRNAs as well as miRNAs. Recently, next-generation sequencing (NGS) miRNA profiles from human HD and neurologically normal control brain tissues were reported. Five consistently upregulated miRNAs affect the expression of genes involved in neuronal differentiation, neurite outgrowth, cell death and survival. We re-analyzed the NGS data publicly available in array express and detected nineteen additional differentially expressed miRNAs. Subsequently, we connected these miRNAs to genes implicated in HD development and network analysis pointed to miRNA-mediated downregulation of twenty-two genes with roles in the pathogenesis as well as treatment of the disease. *In silico* prediction and reporter systems prove that levels of *BDNF*, a central node in the miRNA-mRNA regulatory network, can be post-transcriptionally controlled by upregulated miR-10b-5p and miR-30a-5p. Reduced *BDNF* expression is associated with neuronal dysfunction and death in HD. Moreover, the 3'UTR of *CREB1* harbors a predicted binding site for these two miRNAs. *CREB1* is similarly downregulated in HD and overexpression decreased susceptibility to 3-nitropropionic-induced toxicity in a cell model. In contradiction to these observations, it is presumed that miR-10b-5p upregulation in HD exerts a neuroprotective role in response to the mutation in the huntingtin gene. Therefore, the function of miR-10b-5p and especially its effect on *BDNF* expression in HD requires further academic research.

Keywords: Huntington, miRNA, Sequencing, Post-transcriptional regulation

Introduction

Huntington's disease (HD) is a fatal hereditary neurodegenerative disorder characterized by unwanted choreatic movements, behavioral manifestations and dementia [1]. In the caucasian population HD appears with an incidence of one per 10,000-20,000 per year in middle age (30-50 years) [2]. The disease is caused by a genetic disorder. An elongation of the CAG trinucleotide repeat (36 repeats or more) is observed within the coding region of the huntingtin (*HTT*) gene [3]. This mutation yields a protein with deleterious functions for brain cells and even impairs the

ability of normal *HTT* to exert fundamental molecular activities in the neurons [4]. As a consequence, neurons predominantly degenerate in the brains of affected patients [4]. While the altered biological processes finally leading to neurodegeneration remain poorly understood, changes in messenger RNA (mRNA) expression point to transcriptional dysregulation as a central mechanism [5]. Beside deregulation of mRNAs, also differential expression of microRNAs (miRNAs) has been linked to HD [6]. MiRNAs are a class of small non-coding RNAs (sncRNAs) that can repress gene expression through translational repression or mRNA deadenylation and decay by base pairing to partially complementary sites [7]. Recent research has examined the role of miRNAs in HD using next generation sequencing (NGS) and identified between

Correspondence: s.mueller@bio.uni-frankfurt.de
Molecular BioSciences, University of Frankfurt, Marie-Curie-Str.9, 60439
Frankfurt a.M., Germany

five and 85 deregulated miRNAs [8,9]. Hoss and colleagues [9] related five upregulated miRNAs (miR-10b-5p, miR-196a-5p, miR-196b-5p, miR-615-3p and miR-1247-5p) located in the HOX gene cluster to HD pathogenesis. Nevertheless, target and differential expression analysis with strict parameters revealed only one validated, down-regulated target gene (*KRT5*) of these miRNAs. Therefore, their function in HD pathogenesis mostly remains unclear.

In order to shed light on the consequences of miRNA deregulation in HD we used omiRas [10] to re-analyze the dataset of Hoss and co-workers consisting of small RNA-Sequencing (sRNA-Seq) libraries derived from twelve HD and nine unaffected control brain tissue samples in FASTQ format. In extension to the five miRNAs identified by Hoss and colleagues we detected nineteen additional miRNAs as differentially expressed. Furthermore, we assigned functions to differentially expressed miRNAs via the interaction tool of omiRas. Analysis revealed

BDNF as a validated target of two upregulated miRNAs (miR-10b and miR-30a), *CREB1* is predicted to be post-transcriptionally controlled by the same two miRNAs. The potential miRNA-mediated downregulation of several major player genes in HD pathogenesis underlines the feasibility of miRNAs as therapeutic targets in HD.

Materials and methods

Dataset collection and preprocessing

A publicly available sRNA-Seq expression dataset of twelve HD and nine control brain samples from the prefrontal cortex was downloaded from Array Express (E-MTAB-2206) in FASTQ format. The 3' sequencing adapter (TCGTATGCCGTCTTCTGCTTGAAA) was removed from the reads with cutadapt [11]. Subsequently, low quality stretches below a SANGER quality score of 20 were additionally trimmed from each end of the reads (-q 20). Only reads with a minimum length of fifteen

Table 1 Deregulated miRNAs in HD

miRNA	NEV Control	NEV HD	foldChange	FDR	Other studies
miR-196a-5p	0.00	19.01	Inf	1.4E-010	[9,14]
miR-891a	48.39	101.39	2.10	0.00001	-
miR-10b-5p	1011.52	30689.38	30.34	0.00003	[9]
miR-4645-3p	3.66	9.44	2.58	0.0001	-
miR-1247-5p	135.61	309.04	2.28	0.0004	[9]
miR-10b-3p	0.00	5.28	Inf	0.0026	-
miR-363-3p	2239.63	3274.71	1.46	0.0033	[8]
miR-30a-3p	5223.90	6943.31	1.33	0.0048	[8]
miR-125b-2-3p	17177.77	20740.93	1.21	0.0052	-
miR-615-3p	0.00	5.45	Inf	0.0065	[9]
miR-196b-5p	1.09	10.17	9.33	0.0134	[9]
miR-127-3p	175251.70	224611.39	1.28	0.0194	-
miR-208b	75.76	112.90	1.49	0.0217	-
miR-302a-5p	2.67	6.93	2.60	0.0451	-
miR-2682-5p	212.76	299.86	1.41	0.0451	-
miR-30a-5p	171969.53	228298.92	1.33	0.0451	[8]
miR-770-5p	333.36	445.55	1.34	0.0451	-
miR-130a-3p	4740.57	6385.28	1.35	0.0451	-
miR-92b-5p	40.62	57.17	1.41	0.0451	-
miR-449a	20.31	32.65	1.61	0.0451	-
miR-3139	8.59	2.27	0.26	0.0031	-
miR-4449	10.95	2.75	0.25	0.0163	-
miR-4521	335.51	168.04	0.50	0.0194	-
miR-138-2-3p	94.11	74.60	0.79	0.0194	-

MiRNAs upregulated in HD brains are indicated by a fold-change > 1, downregulated miRNAs by a fold-change < 1. NEV corresponds to the normalized expression value and FDR is the corrected p-value. Other studies indicates if any study different from this has likewise reported the miRNA deregulation in HD.

products of downregulated genes with 121 protein-protein interactions. Hubs in the network represented by nodes with the most protein-protein interactions are Calmodulin 1 (*CALM1*) with twelve interactions and brain-derived neurotrophic factor (*BDNF*) with nine interactions. The downregulation of mRNAs coding for the proteins in the network is potentially caused by eight miRNAs with predicted binding sites in their 3'UTR. Approximately one third (22) of all mRNAs are predicted targets of miRNAs, four genes can be post-transcriptionally controlled by more than one miRNA (*BDNF*, *CALM1*, *CNRI*, *CREB1*). The hub genes *BDNF* and *CALM1* harbor a binding site for miR-10b-5p, 196a-5p, 196b-5p and 30a-5p in their 3'UTR. *CREB1* and *BDNF* are predicted targets of miR-10b and miR-30a, whereas the regulation of *BDNF* has recently been experimentally verified in the prefrontal cortex [15,16].

Discussion

We extend the report of Hoss and co-workers based on NGS miRNA expression profiles of twelve HD and nine healthy control brain samples. Re-analysis of the dataset reveals 24 differentially expressed miRNAs in HD, 20 of these up- and four downregulated. Regulatory network analysis comprising genes involved in HD pathogenesis with decreased expression underlines the role of the most significantly upregulated miRNA, miR-10b-5p, that targets *BDNF* and *CREB1*.

BDNF is a secreted neurotrophic factor, which represent a class of molecules that contribute essentially to the survival of the peripheral and central nervous system, and reduced level of *BDNF* mRNA as well as protein have been found in HD cerebral cortex and striatum [17]. *BDNF* is required in striatal neurons for survival and activity. The largest proportion of striatal *BDNF* is initially produced in the frontal cortex and subsequently transported to the striatum [18]. YAC 128 mice that were transplanted with *BDNF* overexpressing MSCs in the striatum show a significantly reduced amount of neuronal loss [19]. Downregulation of *BDNF* has been directly associated with the mutation of wild-type *HTT* [17]. Our analysis extends the regulatory mechanism leading to *BDNF* downregulation in HD to miR-10b-5p and 30a-5p which are significantly upregulated in HD and have been shown to target the 3'UTR of the *BDNF* transcript [15,16]. Upregulation of *BDNF* levels in the striatum/cortex are a potential therapeutic strategy in HD treatment [18] and our analysis points to an inhibition of miRNAs by antagomiRs to achieve this goal. Mir-10b antagomiRs have *inter alia* been used for therapeutic silencing of miR-10b to inhibit metastasis in a mouse mammary tumor model [20]. In contradiction to these observations, miR-10b-5p expression enhanced the survival of PC12 Q73 cells and its upregulation in HD may

be a neuroprotective response to the *HTT* mutation [9]. Therefore, the role of miR-10b-5p and especially its effect on *BDNF* expression in HD requires further academic research.

CREB1 encodes a transcription factor that is a member of the leucine zipper family of DNA binding proteins. *CREB1* induces transcription of genes in response to hormonal stimulation of the cAMP pathway. Members of the *CREB* family are essential for the maintenance of cell viability in various tissues and stages of development [21]. Reduced *CREB1* expression has been reported in HD and mutant *Htt* represses *CREB1* expression by a direct interaction with the *CREB*-binding protein [22]. Lack of *CREB1* expression during development of the central nervous system leads to substantial apoptosis of postmitotic neurons [21]. The *CREB* signaling pathway has been suggested for pharmacological intervention in neurodegenerative disorders like HD [21]. The 3'UTR of *CREB1* harbors predicted binding sites of miR-10b-5p, 30a-5p and 196a-5p, which makes antagomiRs a potential approach for intervention in *CREB* signalling. Nevertheless, these interactions lack experimental validation and form a basis for further research.

Taken together our analysis underlines the role of miRNAs in HD pathogenesis. The regulatory network of deregulated genes and miRNAs may now spur further research in the field of HD. We provide a set of miRNA-mRNA interactions that currently lack experimental validation and point to miRNAs that are potential targets for treatment with antagomiRs. The validity of the predicted interactions between downregulated genes and upregulated miRNAs is underlined by the recent validation of four interactions in the network (miR-10b-5p-*BDNF*, miR-30a-5p-*BDNF*, miR-30a-5p-*AP2A1*, miR-30a-5p-*PPP3CA* [23]).

Competing interests

The author declares that they have no competing interests.

Acknowledgements

This work was supported by the Bundesministerium für Bildung und Forschung (BMBF) (Grant nos. FKZ0316043 and FKZ031A104A). The author thanks Professor Günter Kahl and Professor Ina Koch as well as Dr. Börn Rotter and Dr. Peter Winter for their advice and assistance.

Received: 29 June 2014 Accepted: 13 August 2014

Published: 18 August 2014

References

1. Pringsheim T, Wiltshire K, Day L, Dykeman J, Steeves T, Jette N: **The incidence and prevalence of Huntington's disease: A systematic review and meta-analysis.** *Mov Disord* 2012, **27**(9):1083–1091.
2. Roos RA: **Huntington's disease: a clinical review.** *Orphanet J Rare Dis* 2010, **5**:40.
3. MacDonald ME, Ambrose CM, Duyao MP, Myers RH, Lin C, Srinidhi L, Barnes G, Taylor SA, James M, Groot N, MacFarlane H, Jenkins B, Anderson MA, Wexler NS, Gusella JF, Bates GP, Baxendale S, Hummerich H, Kirby S, North M, Youngman S, Mott R, Zehetner G, Sedlacek Z, Poustka A, Frischauf A-M, Lehrach H: **A novel gene containing a trinucleotide**

- repeat that is expanded and unstable on Huntington's disease chromosomes. *Cell* 1993, **72**(6):971–983.**
4. Graham RK, Deng Y, Slow EJ, Haigh B, Bissada N, Lu G, Pearson J, Shehadeh J, Bertram L, Murphy Z, Warby SC, Doty CN, Roy S, Wellington CL, Leavitt BR, Raymond LA, Nicholson DW, Hayden MR: **Cleavage at the caspase-6 site is required for neuronal dysfunction and degeneration due to mutant huntingtin.** *Cell* 2006, **125**(6):1179–1191.
 5. Hodges A, Strand AD, Aragaki AK, Kuhn A, Sengstag T, Hughes G, Elliston LA, Hartog C, Goldstein DR, Thu D, Hollingsworth ZR, Collin F, Synek B, Holmans PA, Young AB, Wexler NS, Delorenzi M, Kooperberg C, Augood SJ, Faull RLM, Olson JM, Jones L, Luthi-Carter R: **Regional and cellular gene expression changes in human Huntington's disease brain.** *Hum Mol Genet* 2006, **15**(6):965–977.
 6. Junn E, Mouradian MM: **MicroRNAs in neurodegenerative diseases and their therapeutic potential.** *Pharmacol Ther* 2012, **133**(2):142–150.
 7. Wu L, Fan J, Belasco JG: **MicroRNAs direct rapid deadenylation of mRNA.** *Proc Natl Acad Sci U S A* 2006, **103**(11):4034–4039.
 8. Martí E, Pantano L, Bañez-Coronel M, Llorens F, Miñones-Moyano E, Porta S, Sumoy L, Ferrer I, Estivill X: **A myriad of miRNA variants in control and Huntington's disease brain regions detected by massively parallel sequencing.** *Nucleic Acids Res* 2010, **38**(20):7219–7235.
 9. Hoss AG, Kartha VK, Dong X, Latourelle JC, Dumitriu A, Hadzi TC, MacDonald ME, Gusella JF, Akbarian S, Chen J-F, Weng Z, Myers RH: **MicroRNAs located in the Hox gene clusters are implicated in huntington's disease pathogenesis.** *PLoS Genet* 2014, **10**(2):e1004188.
 10. Müller S, Rycak L, Winter P, Kahl G, Koch I, Rotter B: **omiRas: a Web server for differential expression analysis of miRNAs derived from small RNA-Seq data.** *Bioinformatics* 2013, **29**(20):2651–2652.
 11. Martin M: **Cutadapt removes adapter sequences from high-throughput sequencing reads.** *EMBnet J* 2011, **17**:10.
 12. Kalathur RKR, Hernández-Prieto MA, Futschik ME: **Huntington's disease and its therapeutic target genes: a global functional profile based on the HD research crossroads database.** *BMC Neurol* 2012, **12**:47.
 13. Anders S, Huber W: **Differential expression analysis for sequence count data.** *Genome Biol* 2010, **11**(10):R106.
 14. Packer AN, Xing Y, Harper SQ, Jones L, Davidson BL: **The bifunctional microRNA miR-9/miR-9* regulates REST and CoREST and is downregulated in Huntington's disease.** *J Neurosci* 2008, **28**(53):14341–14346.
 15. Varendi K, Kumar A, Härma MA, Andressoo JO: **miR-1, miR-10b, miR-155, and miR-191 are novel regulators of BDNF.** *Cell Mol Life Sci* 2014, **2014**:1–14.
 16. Mellios N, Huang HS, Grigorenko A, Rogaev E, Akbarian S: **A set of differentially expressed miRNAs, including miR-30a-5p, act as post-transcriptional inhibitors of BDNF in prefrontal cortex.** *Hum Mol Genet* 2008, **17**(19):3030–3042.
 17. Zuccato C, Ciammola A, Rigamonti D, Leavitt BR, Goffredo D, Conti L, MacDonald ME, Friedlander RM, Silani V, Hayden MR, Timmusk T, Sipione S, Cattaneo E: **Loss of huntingtin-mediated BDNF gene transcription in Huntington's disease.** *Science* 2001, **293**(5529):493–498.
 18. Xie Y, Hayden MR, Xu B: **BDNF overexpression in the forebrain rescues Huntington's disease phenotypes in YAC128 mice.** *J Neurosci* 2010, **30**(44):14708–14718.
 19. Dey ND, Bombard MC, Roland BP, Davidson S, Lu M, Rossignol J, Sandstrom MI, Skeel RL, Lescaudron L, Dunbar GL: **Genetically engineered mesenchymal stem cells reduce behavioral deficits in the YAC 128 mouse model of Huntington's disease.** *Behav Brain Res* 2010, **214**(2):193–200.
 20. Ma L, Reinhardt F, Pan E, Soutschek J, Bhat B, Marcusson EG, Teruya-Feldstein J, Bell GW, Weinberg RA: **Therapeutic silencing of miR-10b inhibits metastasis in a mouse mammary tumor model.** *Nat Biotechnol* 2010, **28**(4):341–347.
 21. Mantamadiotis T, Lemberger T, Bleckmann SC, Kern H, Kretz O, Villalba AM, Tronche F, Kellendonk C, Gau D, Kapfhammer J, Otto C, Schmid W, Schütz G: **Disruption of CREB function in brain leads to neurodegeneration.** *Nat Genet* 2002, **31**:47–54.
 22. Chaturvedi RK, Hennessey T, Johri A, Tiwari SK, Mishra D, Agarwal S, Kim YS, Beal MF: **Transducer of regulated CREB-binding proteins (TORCs) transcription and function is impaired in Huntington's disease.** *Hum Mol Genet* 2012, **21**(15):3474–3488.
 23. Vergoulis T, Vlachos IS, Alexiou P, Georgakilas G, Maragkakis M, Reczko M, Gerangelos S, Koziris N, Dalamagas T, Hatzigeorgiou AG: **TarBase 6.0: capturing the exponential growth of miRNA targets with experimental support.** *Nucleic Acids Res* 2012, **40**(D1):D222–D229.

doi:10.1186/2047-9158-3-17

Cite this article as: Müller: *In silico* analysis of regulatory networks underlines the role of miR-10b-5p and its target BDNF in huntington's disease. *Translational Neurodegeneration* 2014 **3**:17.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

